



Europäisches Patentamt  
European Patent Office  
Office européen des brevets



(11) EP 0 897 005 A1

(12) EUROPEAN PATENT APPLICATION

(43) Date of publication:  
17.02.1999 Bulletin 1999/07

(51) Int Cl.<sup>6</sup>: C12N 15/52, C12N 15/60,  
C12N 9/88, C12N 15/74,  
C12N 1/21

(21) Application number: 98302554.5

(22) Date of filing: 01.04.1998

(84) Designated Contracting States:  
AT BE CH DE DK ES FI FR GB GR IE IT LI LU MC  
NL PT SE  
Designated Extension States:  
AL LT LV MK RO SI

(30) Priority: 01.04.1997 JP 82965/97

(71) Applicants:  
• Japan Science and Technology Corporation  
Kawaguchi-shi, Saitama 332-0012 (JP)  
• THE INSTITUTE OF PHYSICAL & CHEMICAL  
RESEARCH  
Wako-shi, Saitama 351-0198 (JP)

(72) Inventors:  
• Dol, Yoshiharu  
Wako-shi, Saitama 351-0198 (JP)  
• Fukui, Toshiaki  
Wako-shi, Saitama 351-0198 (JP)  
• Matsusaki, Hiromi  
Wako-shi, Saitama 351-0115 (JP)

(74) Representative: Nachshen, Nell Jacob et al  
D Young & Co  
21 New Fetter Lane  
London EC4A 1DA (GB)

Remarks:

The applicant has subsequently filed a sequence  
listing and declared, that it includes no new matter.

(54) Polyester synthase and a gene coding for the same

(57) The present invention relates to a polypeptide  
comprising the amino acid sequence of SEQ ID NO:1  
or a sequence where in said amino acid sequence, one  
or more amino acids are deleted, replaced or added,

said polypeptide having polyester synthase activity; a  
polyester synthase gene comprising DNA coding for  
said polypeptide; a recombinant vector comprising the  
gene; and a transformant transformed with the recom-  
binant vector.

EP 0 897 005 A1

## Description

## Field of the Invention

5 [0001] The present invention relates to polyester synthase, a gene coding for the enzyme, a recombinant vector containing the gene, a transformant transformed with the vector, and a process for producing polyester synthase by use of the transformant.

## Background of the Invention

10 [0002] Polyesters (e.g. poly-3-hydroxyalkanoic acid) biosynthesized by microorganisms are biodegradable plastics with thermoplasticity ranging widely from rigid matter to viscoelastic rubber.

[0003] Poly-3-hydroxybutanoic acid (P(3HB)) is a typical polyester consisting of C4 monomer units, but it is a rigid and brittle polymeric material, so its application is limited. Accordingly, various polyesters such as P(3HB-co-3HV) having (P(3HB)) copolymerized with a C5 monomer unit (3HV) by adding propionic acid etc. to the medium have been prepared and examined to alter the physical properties of the polyester. On the other hand, polyesters consisting of at least C6 monomer units are soft polymeric materials having plasticity.

[0004] Polyester-synthesizing microorganisms are roughly divided into 2 groups, that is, those synthesizing polyesters with C3-5 monomer units and those synthesizing polyesters with C6-14 monomer units. The former microorganisms possess a polyester synthase using C3-5 monomer units as the substrate, while the latter microorganisms possess a polyester synthase using C6-14 monomer units as the substrate. Therefore, polyesters with different properties are synthesized by the respective microorganisms.

[0005] However, the respective polyesters from such known microorganisms are different in substrate specificity, so with one kind of enzyme given, polyesters (copolymers) having various monomer unit compositions adapted to the object of use are difficult to synthesize.

## Summary of the Invention

[0006] The object of the present invention is to provide a polyester synthase preferably having specificity for monomer units having a wide range of carbon atoms as the substrate, a gene coding for the enzyme, a recombinant vector containing the gene, a transformant transformed with the vector, and a process for producing the polyester synthase by use of the transformant.

[0007] As a result of their eager research, the present inventors succeeded in cloning a polyester synthase gene from a microorganism belonging to the genus Pseudomonas isolated from soil, to arrive at the completion of the present invention.

[0008] That is, the present invention relates to a polypeptide comprising the amino acid sequence of SEQ ID NO:1, analogs, variants or fragments thereof, for example where one or more amino acids are deleted, replaced or added, said polypeptide having polyester synthase activity.

[0009] Further, the present invention relates to a polyester synthase gene comprising DNA coding for said polypeptide or variants or fragments thereof. The DNA coding for the protein with polyester synthase activity includes e.g. that of SEQ ID NO:2.

[0010] Further, the present invention relates to a polyester synthase gene comprising the nucleotide sequence of SEQ ID NO:3.

[0011] Further, the present invention relates to a recombinant vector comprising the polyester synthase gene.

[0012] Further, the present invention relates to a transformant transformed with said recombinant vector.

[0013] Further, the present invention relates to a process for producing polyester synthase wherein said transformant is cultured in a medium and polyester synthase is recovered from the resulting culture.

## Detailed Description of the Invention

50 [0014] Hereinafter, the present invention is described in detail.

## (1) Cloning of the polyester synthase gene

55 [0015] The polyester synthase gene of the present invention may be isolated from a microorganism belonging to the genus Pseudomonas.

[0016] First, genomic DNA is isolated from a strain having the polyester synthase gene. Such a strain includes e.g. Pseudomonas sp. Any known methods can be used for preparation of genomic DNA. For example, Pseudomonas sp.

is cultured in a bouillon medium and then its genomic DNA is prepared by the hexadecyl trimethyl ammonium bromide method (Current Protocols in Molecular Biology, vol. 1, page 2.4.3., John Wiley & Sons Inc., 1994).

[0017] The DNA obtained in this manner is partially digested with a suitable restriction enzyme (e.g. *Sau3AI*, *BamHI*, *BglII* etc.). It is then ligated into a vector dephosphorylated by treatment with alkaline phosphatase after cleavage with a restriction enzyme (e.g. *BamHI*, *BglII* etc.) to prepare a library.

[0018] Phage or plasmid capable of autonomously replicating in host microorganisms is used as the vector. The phage vector includes e.g. *EMBL3*, *M13*,  $\lambda$ gt11 etc., and the plasmid vector includes e.g. *pBR322*, *pUC18*, and *pBlue-script II* (Stratagene). Vectors capable of autonomously replicating in 2 or more host cells such as *E. coli* and *Bacillus brevis*, as well as various shuttle vectors, can also be used. Such vectors are also cleaved with said restriction enzymes so that their fragment can be obtained.

[0019] Conventional DNA ligase is used to ligate the resulting DNA fragment into the vector fragment. The DNA fragment and the vector fragment are annealed and then ligated to produce a recombinant vector.

[0020] To introduce the recombinant vector into a host microorganism, any known methods can be used. For example, if the host microorganism is *E. coli*, the calcium chloride method (Lederberg, E.M. et al., *J. Bacteriol.* **119**, 1072 (1974)) and the electroporation method (Current Protocols in Molecular Biology, vol. 1, page 1.8.4 (1994)) can be used. If phage DNA is used, the *in vitro* packaging method (Current Protocols in Molecular Biology, vol. 1, page 5.7.1 (1994)) etc. can be adopted. In the present invention, an *in vitro* packaging kit (*Gigapack II*, produced by Stratagene etc.) may be used.

[0021] To obtain a DNA fragment containing the polyester synthase gene derived from *Pseudomonas* sp., a probe is then prepared. The amino acid sequences of some polyester synthases have already been known (Peoples, O. P. and Sinskey, A. J., *J. Biol. Chem.*, **264**, 15293 (1989); Huisman, G. W. et al., *J. Biol. Chem.*, **266**, 2191 (1991); Pieper, U. et al., *FEMS Microbiol. Lett.*, **96**, 73 (1992); Timm, A. and Steinbuchel, A., *Eur. J. Biochem.*, **209**, 15 (1992), etc.). Well-conserved regions are selected from these amino acid sequences, and nucleotide sequences coding for them are estimated to design oligonucleotides. Examples of such oligonucleotides include, but are not limited to, the sequence 5'-CC(G/C)CAGATCAACAAGTT(C/T)TA(C/G)GAC-3' (SEQ ID NO:4) reported by Timm, A. and Steinbuchel, A., *Eur. J. Biochem.*, **209**, 15 (1992).

[0022] Then, this synthetic oligonucleotide is labeled with a suitable reagent and used for colony hybridization of the above genomic DNA library (Current Protocols in Molecular Biology, vol. 1, page 6.0.3 (1994)).

[0023] The *E. coli* is screened by colony hybridization, and a plasmid is recovered from it using the alkaline method (Current Protocols in Molecular Biology, vol. 1, page 1.6.1 (1994)), whereby a DNA fragment containing the polyester synthase gene is obtained. The nucleotide sequence of this DNA fragment can be determined in e.g. an automatic nucleotide sequence analyzer such as 373A DNA sequencer (Applied Biosystems) using a known method such as the Sanger method (Molecular Cloning, vol. 2, page 13.3 (1989)).

[0024] After the nucleotide sequence was determined by the means described above, the gene of the present invention can be obtained by chemical synthesis or the PCR technique using genomic DNA as a template, or by hybridization using a DNA fragment having said nucleotide sequence as a probe.

## (2) Preparation of transformant

[0025] The transformant of the present invention is obtained by introducing the recombinant vector of the present invention into a host compatible with the expression vector used in constructing said recombinant vector.

[0026] The host is not particularly limited insofar as it can express the target gene. Examples are bacteria such as microorganisms belonging to the genus *Alcaligenes*, microorganisms belonging to the genus *Bacillus*, bacteria such as *E. coli*, yeasts such as the genera *Saccharomyces*, *Candida* etc., and animal cells such as COS cells, CHO cells etc.

[0027] If microorganisms belonging to the genus *Alcaligenes* or bacteria such as *E. coli* are used as the host, the recombinant DNA of the present invention is preferably constituted such that it contains a promoter, the DNA of the present invention, and a transcription termination sequence so as to be capable of autonomous replication in the host. The expression vector includes *pLA2917* (ATCC 37355) containing replication origin *RK2* and *pJRD215* (ATCC 37533) containing replication origin *RSF1010*, which are replicated and maintained in a broad range of hosts.

[0028] The promoter may be any one if it can be expressed in the host. Examples are promoters derived from *E. coli*, phage etc., such as *trp* promoter, *lac* promoter, *PL* promoter, *PR* promoter and *T7* promoter. The method of introducing the recombinant DNA into bacteria includes e.g. a method using calcium ions (Current Protocols in Molecular Biology, vol. 1, page 1.8.1 (1994)) and the electroporation method (Current Protocols in Molecular Biology, vol. 1, page 1.8.4 (1994)).

[0029] If yeast is used as the host, expression vectors such as *YEpl3*, *YCp50* etc. are used. The promoter includes e.g. *gal 1* promoter, *gal 10* promoter etc. To method of introducing the recombinant DNA into yeast includes e.g. the electroporation method (Methods. Enzymol., **194**, 182-187 (1990)), the spheroplast method (Proc. Natl. Acad. Sci. USA, **84**, 1929-1933 (1978)), the lithium acetate method (*J. Bacteriol.*, **153**, 163-168 (1983)) etc.

[0030] If animal cells are used as the host, expression vectors such as pcDNA1, pcDNA1/Amp (produced by Invitro-gene) etc. are used. The method of introducing the recombinant DNA into animal cells includes e.g. the electroporation method, potassium phosphate method etc.

### 5 (3) Production of polyester synthase

[0031] Production of the polyester synthase of the present invention is carried out by culturing the transformant of the present invention in a medium, forming and accumulating the polyester synthase of the present invention in the culture (the cultured microorganism or the culture supernatant) and recovering the polyester synthase from the culture.

10 [0032] A conventional method used for culturing the host is also used to culture the transformant of the present invention.

[0033] The medium for the transformant prepared from bacteria such as *E. coli* etc. as the host includes complete medium or synthetic medium, e.g. LB medium, M9 medium etc. The transformant is aerobically cultured at a temperature ranging from 25 to 37 °C for 12 to 48 hours so that the polyester synthase is accumulated in the microorganism and then recovered.

15 [0034] The carbon source is essential for the growth of the microorganism and includes e.g. carbohydrates such as glucose, fructose, sucrose, maltose etc.

[0035] The nitrogen source includes e.g. ammonia, ammonium salts such as ammonium chloride, ammonium sulfate, ammonium phosphate etc., peptone, meat extract, yeast extract, corn steep liquor etc. The inorganic matter includes e.g. monopotassium phosphate, dipotassium phosphate, magnesium phosphate, magnesium sulfate, sodium chloride etc.

[0036] Culture is carried out usually under aerobic conditions with shaking at 25 to 37 °C for more than 2 hours after expression is induced. During culture, antibiotics such as ampicillin, kanamycin, ampicillin, tetracycline etc. may be added to the culture.

25 [0037] To culture the microorganism transformed with the expression vector using an inducible promoter, its inducer can also be added to the medium. For example, isopropyl- $\beta$ -D-thiogalactopyranoside (IPTG), indoleacrylic acid (IAA) etc. can be added to the medium.

[0038] To culture the transformant from animal cells as the host, use is made of a medium such as RPMI-1640 or DMEM which may be supplemented with fetal bovine serum. Culture is carried out usually in 5 % CO<sub>2</sub> at 30 to 37°C for 1 to 7 days. During culture, antibiotics such as kanamycin, penicillin etc. may be added to the medium.

30 [0039] Purification of the polyester synthase can be performed by recovering the resulting culture by centrifugation (after disruption in the case of cells) and subjecting it to affinity chromatography, cation or anion exchange chromatography or gel filtration or to a suitable combination thereof.

[0040] Whether the resulting purified substance is the desired enzyme is confirmed by conventional methods such as SDS polyacrylamide gel electrophoresis, Western blotting etc.

### Examples

40 [0041] Hereinafter, the present invention is described in more detail with reference to the Examples which however are not intended to limit the scope of the present invention.

#### (1) Cloning of the polyester synthase gene from *Pseudomonas* sp.

[0042] First, a genomic DNA library of *Pseudomonas* sp. was prepared.

45 [0043] *Pseudomonas* sp. JCM 10015 was cultured overnight in 100 ml bouillon medium (1 % meat extract, 1 % peptone, 0.5 % sodium chloride, pH 7.2) at 30 °C and then genomic DNA was obtained from the microorganism using the hexadecyl trimethyl ammonium bromide method (Current Protocols in Molecular Biology, vol. 1, page 2.4.3 (1994), John Wiley & Sons Inc.).

[0044] The resulting genomic DNA was partially digested with restriction enzyme Sau3AI. The vector plasmid used was cosmid vector pLA2917 (ATCC 37355). This plasmid was cleaved with restriction enzyme BglII and dephosphorylated (Molecular Cloning, vol. 1, page 5.7.2 (1989), Cold Spring Harbor Laboratory) and then ligated into the partially digested genomic DNA fragment by use of DNA ligase.

50 [0045] *E. coli* S17-1 was transformed with this ligated DNA fragment by the *in vitro* packaging method (Current Protocols in Molecular Biology, vol. 1, page 5.7.2 (1994)) whereby a genomic DNA library from *Pseudomonas* sp. was obtained.

[0046] To obtain a DNA fragment containing the polyester synthase gene from *Pseudomonas* sp., a probe was then prepared. An oligonucleotide consisting of the sequence 5'-CC(G/C)CAGATCAACAAGTT(C/T)TA(C/G)GAC-3' (SEQ ID NO:4) reported by Timm, A. and Steinbuechel, A., Eur. J. Biochem., 209, 15 (1992) was synthesized. This oligonu-

cleotide was labeled with digoxigenin using a DIG DNA labeling kit (Boehringer Mannheim) and used as a probe.

[0047] Using the probe thus obtained, *E. coli* carrying a plasmid containing the polyester synthase gene was isolated by colony hybridization from the genomic DNA library from *Pseudomonas* sp.

[0048] When *Alcaligenes eutrophus* PHB-4 (DSM541) and *Pseudomonas putida* GPp104 (both of which are strains deficient in an ability to produce polyester) were transformed by the conjugation transfer method with the plasmid containing the polyester synthase gene, both the strains had a reverse ability to produce polyester and showed complementarity.

[0049] By recovering the plasmid from the *E. coli*, a DNA fragment containing the polyester synthase gene was obtained.

[0050] The nucleotide sequence of a PstI-XbaI fragment from this fragment was determined by the Sanger method.

[0051] As a result, the nucleotide sequence of the 1.8 kbp fragment shown in SEQ ID NO:3 was determined.

[0052] By further examining homology to this nucleotide sequence, the polyester synthase gene containing the nucleotide sequence (1680 bp) of SEQ ID NO:2 could be identified in this 1.8 kbp nucleotide sequence. The amino acid sequence encoded by SEQ ID NO:2 is shown in SEQ ID NO:1.

[0053] It should be understood that insofar as a protein containing the amino acid sequence of SEQ ID NO:1 or a sequence where in said amino acid sequence, one or more amino acids are deleted, replaced or added has polyester synthase activity, the gene (SEQ ID NO:2 or 3) containing DNA coding for said protein falls under the scope of the polyester synthase gene of the present invention.

[0054] Mutations such as deletion, replacement, addition etc. can be induced in the amino acid sequence or nucleotide sequence by the known site-directed mutagenesis method (e.g. Transformer™ Site-Directed Mutagenesis Kit available from Toyobo).

## (2) Preparation of *E. coli* transformant.

[0055] The 1.8 kb PstI-XbaI fragment containing the polyester synthase gene was ligated into the XbaI, PstI site of plasmid vector pBluescript II KS+. The resulting recombinant vector was transformed by the calcium chloride method into *Escherichia coli* DH5α. The resulting transformant was designated *Escherichia coli* PX18. By extracting the plasmid from this transformant, the 1.8 kb PstI-XbaI fragment containing the polyester synthase gene can be easily obtained. *Escherichia coli* PX18 has been deposited as FERM BP-6297 with the National Institute of Bioscience and Human Technology, Agency of Industrial Science and Technology, Japan (1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken 305-8566, Japan).

[0056] According to the present invention, there are provided a gene coding for polyester synthase, a recombinant vector containing the gene, and a transformant transformed with the vector. The gene of the present invention codes for a polyester synthase using monomers having a wide range of carbon atoms as the substrate, so it is useful in preparing copolymer polyesters having various physical properties.

[0057] The term "analog" as used herein, in relation to proteins or polypeptides of the present invention includes any peptidomimetic, that is, a chemical compound that possesses polyester synthase activity in a similar manner to the parent protein.

[0058] The term "variant" as used herein, in relation to proteins or polypeptides of the present invention includes any substitution of, variation of, modification of, replacement of, deletion of or addition of one (or more) amino acid residues from or to the sequence providing that the resultant protein or polypeptide possesses polyester synthase activity.

[0059] The term "fragment" as used herein, in relation to proteins or polypeptides of the present invention includes any shorter forms of the polypeptides of the present invention that possesses polyester synthase activity in a similar manner to the parent protein.

[0060] The terms "variants" or "fragments" as used in relation to the DNA molecules of the present invention, have the meanings as given above (as appropriate to DNA) given that the DNA variant, fragment or derivative encodes a polypeptide that possesses polyester synthase activity in a similar manner to the parent protein.

Annex to the description

[0061]

5

## SEQUENCE LISTING

10

## (2) INFORMATION FOR SEQ ID NO: 1:

15

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 559 amino acids

(B) TYPE: amino acid

20

(C) STRANDEDNESS:

(D) TOPOLOGY: linear

25

## (ii) MOLECULE TYPE: protein

30

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

35

Met Ser Asn Lys Asn Ser Asp Asp Leu Asn Arg Gln Ala Ser Glu Asn

1 5 10 15

Thr Leu Gly Leu Asn Pro Val Ile Gly Leu Arg Gly Lys Asp Leu Leu

20 25 30

40

Thr Ser Ala Arg Met Val Leu Thr Gln Ala Ile Lys Gln Pro Ile His

35 40 45

Ser Val Lys His Val Ala His Phe Gly Ile Glu Leu Lys Asn Val Met

45

50 55 60

Phe Gly Lys Ser Lys Leu Gln Pro Glu Ser Asp Asp Arg Arg Phe Asn

65 70 75 80

50

Asp Pro Ala Trp Ser Gln Asn Pro Leu Tyr Lys Arg Tyr Leu Gln Thr

85 90 95

Tyr Leu Ala Trp Arg Lys Glu Leu His Asp Trp Ile Gly Asn Ser Lys

55

100 105 110

Leu Ser Glu Gln Asp Ile Asn Arg Ala His Phe Val Ile Thr Leu Met  
 115 120 125  
 5 Thr Glu Ala Met Ala Pro Thr Asn Ser Ala Ala Asn Pro Ala Ala Val  
 130 135 140  
 10 Lys Arg Phe Phe Glu Thr Gly Gly Lys Ser Leu Leu Asp Gly Leu Thr  
 145 150 155 160  
 15 His Leu Ala Lys Asp Leu Val Asn Asn Gly Gly Met Pro Ser Gln Val  
 165 170 175  
 Asp Met Gly Ala Phe Glu Val Gly Lys Ser Leu Gly Thr Thr Glu Gly  
 180 185 190  
 20 Ala Val Val Phe Arg Asn Asp Val Leu Glu Leu Ile Gln Tyr Arg Pro  
 195 200 205  
 25 Thr Thr Glu Gln Val His Glu Arg Pro Leu Leu Val Val Pro Pro Gln  
 210 215 220  
 Ile Asn Lys Phe Tyr Val Phe Asp Leu Ser Pro Asp Lys Ser Leu Ala  
 225 230 235 240  
 30 Arg Phe Cys Leu Ser Asn Asn Gln Gln Thr Phe Ile Val Ser Trp Arg  
 245 250 255  
 35 Asn Pro Thr Lys Ala Gln Arg Glu Trp Gly Leu Ser Thr Tyr Ile Asp  
 260 265 270  
 40 Ala Leu Lys Glu Ala Val Asp Val Val Ser Ala Ile Thr Gly Ser Lys  
 275 280 285  
 Asp Ile Asn Met Leu Gly Ala Cys Ser Gly Gly Ile Thr Cys Thr Ala  
 290 295 300  
 45 Leu Leu Gly His Tyr Ala Ala Leu Gly Glu Lys Lys Val Asn Ala Leu  
 305 310 315 320  
 50 Thr Leu Leu Val Ser Val Leu Asp Thr Thr Leu Asp Ser Gln Val Ala  
 325 330 335  
 55 Leu Phe Val Asp Glu Lys Thr Leu Glu Ala Ala Lys Arg His Ser Tyr

340 345 350  
 Gln Ala Gly Val Leu Glu Gly Arg Asp Met Ala Lys Val Phe Ala Trp  
 5 355 360 365  
 Met Arg Pro Asn Asp Leu Ile Trp Asn Tyr Trp Val Asn Asn Tyr Leu  
 10 370 375 380  
 Leu Gly Asn Glu Pro Pro Val Phe Asp Ile Leu Phe Trp Asn Asn Asp  
 15 385 390 395 400  
 Thr Thr Arg Leu Pro Ala Ala Phe His Gly Asp Leu Ile Glu Met Phe  
 405 410 415  
 20 Lys Asn Asn Pro Leu Val Arg Ala Asn Ala Leu Glu Val Ser Gly Thr  
 420 425 430  
 25 Pro Ile Asp Leu Lys Gln Val Thr Ala Asp Ile Tyr Ser Leu Ala Gly  
 435 440 445  
 Thr Asn Asp His Ile Thr Pro Trp Lys Ser Cys Tyr Lys Ser Ala Gln  
 30 450 455 460  
 Leu Phe Gly Gly Lys Val Glu Phe Val Leu Ser Ser Ser Gly His Ile  
 35 465 470 475 480  
 Gln Ser Ile Leu Asn Pro Pro Gly Asn Pro Lys Ser Arg Tyr Met Thr  
 485 490 495  
 40 Ser Thr Asp Met Pro Ala Thr Ala Asn Glu Trp Gln Glu Asn Ser Thr  
 500 505 510  
 45 Lys His Thr Asp Ser Trp Trp Leu His Trp Gln Ala Trp Gln Ala Glu  
 515 520 525  
 Arg Ser Gly Lys Leu Lys Lys Ser Pro Thr Ser Leu Gly Asn Lys Ala  
 50 530 535 540  
 Tyr Pro Ser Gly Glu Ala Ala Pro Gly Thr Tyr Val His Glu Arg  
 55 545 550 555



## (2) INFORMATION FOR SEQ ID NO: 2:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1680 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: DNA (genomic)

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

25 ATGAGTAACA AGAATAGOGA TGACTTGAAT CGTCAAGCCT CGGAAAACAC CTTGGGGCTT 60  
 AACCCCTGTCA TCGGCCTGCG TGGAAAAGAT CTGCTGACTT CTGCCCGAAT GGTTTTAACC 120  
 CAAGCCATCA AACAACCCAT TCACAGCGTC AAGCACGTCG CGCATTTTGG CATCGAGCTG 180  
 30 AAGAACGTGA TGTTTGGCAA ATCGAAGCTG CAACCGGAAA GCGATGACCG TCGTTTCAAC 240  
 GACCCCGCCT GGAGTCAGAA CCCACTCTAC AAACGTTATC TACAAACCTA CCTGGCGTGG 300  
 CGCAAGGAAC TCCACGACTG GATCGGCAAC AGCAAACGTG CCGAACAGGA CATCAATCGC 360  
 35 GCTCACTTCG TGATCACCTT GATGACCGAA GCCATGGCCC CGACCAACAG TGCGGCCAAT 420  
 CCGGCGGCGG TCAAACGCTT CTTGAAACC GCGGTAAAA GCCTGCTCGA CGGCCTCACA 480  
 40 CATCTGGCCA AGGACCTGGT AAACAACGGC GGCATGCCGA GCCAGGTGGA CATGGGCGCT 540  
 TTCGAAGTCG GCAAGAGTCT GGGGACGACT GAAGGTGCAG TGGTTTTCCG CAACGACGTC 600  
 CTCGAATTGA TCCAGTACCG GCCGACCACC GAACAGGTGC ATGAGCGACC GCTGCTGGTG 660  
 45 GTCCACCCGC AGATCAACAA GTTTTATGTG TTTGACCTGA GCGCGGATAA AAGCCTGGCG 720  
 CGCTTCTGCC TGAGCAACAA CCAGCAAACC TTTATCGTCA GCTGGCGCAA CCCGACCAAG 780  
 50 GCCAGCGTG AGTGGGGTCT GTCGACTTAC ATCGATGCGC TCAAAGAAGC CGTCGACGTA 840  
 GTTTCCGCCA TCACCGGCAG CAAAGACATC AACATGCTCG GCGCCTGCTC CCGTGGCATT 900  
 ACCTGCACCG CGCTGCTGGG TCACTACGCC GCTCTCGGG AGAAGAAGGT CAATGCCCTG 960  
 55 ACCCTTTTGG TCAGCGTGCT CGACACCACC CTCGACTCCC AGGTTGCACT GTTCGTCGAT 1020

GAGAAAACCC TGGAAGCTGC CAAGCGTCAC TCGTATCAGG CCGGCGTGCT GGAAGGCCGC 1080  
 GACATGGCCA AAGTCTTCGC CTGGATGCCG CCTAACGACC TGATCTGGAA CTA CTGGGTC 1140  
 5 AACAACTACC TGCTGGGTAA CGAGCCACCG GTCTTCGACA TTCTTTTCTG GAACAACGAC 1200  
 ACCACCCGGT TGCCTGCTGC GTTCCACGGC GATCTGATCG AAATGTTCAA AAATAACCCA 1260  
 10 CTGGTGGCGG CCAATGCACT CGAAGTGAGC GGCACGCCGA TCGACCTCAA ACAGGTCAC 1320  
 GCGGACATCT ACTCCCTGGC CGGCACCAAC GATCACATCA CGCCCTGGAA GTCTTGCTAC 1380  
 AAGTCGGCGC AACTGTTTCGG TGGCAAGGTC GAATTCGTGC TGTCCAGCAG TGGGCATATC 1440  
 15 CAGAGCATTC TGAACCCGCC GGGCAATCCG AAATCACGTT ACATGACCAG CACCGACATG 1500  
 CCAGCCACCG CCAACGAGTG GCAAGAAAAC TCAACCAAGC ACACCGACTC CTGGTGGCTG 1560  
 CACTGGCAGG CCTGGCAGGC CGAGCGCTCG GGCAAACTGA AAAAGTCCCC GACCAGCCTG 1620  
 20 GGCAACAAGG CCTATCCGTC AGGAGAAGCC GCGCCGGGCA CGTATGTGCA TGAACGTAA 1680

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1826 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

CTGCAGTGCT CTCTGAACTA GAAAGCAACG TTGTGCAATT AACGGTCACC CGAGCAGTAG 60  
 TACCTGGCGG TTGCTGTGTG ACTACACAGC TGGTCCCGGT ACTCGTCTCA GGACAATGGA 120  
 GCGTCGTAGA TGAGTAACAA GAATAGCGAT GACTTGAATC GTCAAGCCTC GGAAAACACC 180  
 55 TTGGGGCTTA ACCCTGTCAT CGGCCTGCGT GGAAAAGATC TGCTGACTTC TGCCCGAATG 240

GTTTTAACCC AAGCCATCAA ACAACCCATT CACAGCGTCA AGCACGTCGC GCATTTTGGC 300  
 5 ATCGAGCTGA AGAACGTGAT GTTTGGCAAA TCGAAGCTGC AACCGGAAAG CGATGACCGT 360  
 CGTTTCAACG ACCCGCGCTG GAGTCAGAAC CCACTCTACA AACGTTATCT ACAAACCTAC 420  
 CTGGCGTGGC GCAAGGAACT CCAGGACTGG ATCGGCAACA GCAAAGTGTG CGAACAGGAC 480  
 10 ATCAATCGCG CTCACTTCGT GATCACCCTG ATGACCGAAG CCATGGCCCC GACCAACAGT 540  
 GCGGCCAATC CGGCGGCGGT CAAACGCTTC TTCGAAACCG GCGGTAAAAG CCTGCTCGAC 600  
 15 GGCTCACAC ATCTGGCCAA GGACCTGGTA AACACGGCG GCATGCCGAG CCAGGTGGAC 660  
 ATGGGCGCTT TCGAAGTCGG CAAGAGTCTG GGGACGACTG AAGGTGCAGT GGTTTTCCGC 720  
 AACGACGTCC TCGAATTGAT CCAGTACCGG CCGACCACCG AACAGGTGCA TGAGCGACCG 780  
 20 CTGCTGGTGG TCCCACCGCA GATCAACAAG TTTTATGTGT TTGACCTGAG CCGGATAAA 840  
 AGCCTGGCGC GCTTCTGCCT GAGCAACAAC CAGCAAACCT TTATCGTCAG CTGGCGCAAC 900  
 25 CCGACCAAGG CCCAGCGTGA GTGGGGTCTG TCGACTTACA TCGATGCGCT CAAAGAAGCC 960  
 GTCGACGTAG TTTCCGCCAT CACCGGCAGC AAAGACATCA ACATGCTCGG CGCCTGCTCC 1020  
 GGTGGCATT A CTGCACCGC GCTGCTGGGT CACTACGCCG CTCTCGGCGA GAAGAAGGTC 1080  
 30 AATGCCCTGA CCTTTTGGT CAGCGTGCTC GACACCACCC TCGACTCCCA GGTGCACTG 1140  
 TTCGTGATG AGAAAAOCCT GGAAGCTGCC AAGGTCCTACT CGTATCAGGC CGGCGTGCTG 1200  
 35 GAAGGCCGCG ACATGGOCOA AGTCTTCGCC TGGATGCGCC CTAACGACCT GATCTGGAAC 1260  
 TACTGGGTCA ACAACTACCT GCTGGGTAAAC GAGCCACCGG TCTTCGACAT TCTTTTCTGG 1320  
 AACAAAGACA CCACCCGGTT GCCTGCTGCG TTCCACGGCG ATCTGATCGA AATGTTCAAA 1380  
 40 AATAACCCAC TGGTGCGCGC CAATGCACTC GAAGTGAGCG GCACGCCGAT CGACCTCAAA 1440  
 CAGGTCACTG CCGACATCTA CTCCTGGCC GGCACCAACG ATCACATCAC GCCCTGGAAG 1500  
 TCTTGCTACA AGTCGGCGCA ACTGTTGGT GGCAAGGTCG AATTCGTGCT GTCCAGCAGT 1560  
 45 GGGCATATCC AGAGCATTCT GAACCCGCCG GGCAATCCGA AATCAGTTA CATGACCAGC 1620  
 ACCGACATGC CAGCCACCGC CAACGAGTGG CAAGAAACT CAACCAAGCA CACCGACTCC 1680  
 50 TGGTGGCTGC ACTGGCAGGC CTGGCAGGCC GAGCGCTCGG GCAAAGTAA AAAGTCCCCG 1740  
 ACCAGCCTGG GCAACAAGGC CTATCCGTCA GGAGAAGCCG CGCCGGGCAC GTATGTGCAT 1800  
 GAACGTTAAG TTGTAGGCAG TCTAGA 1826

55

(2) INFORMATION FOR SEQ ID NO: 4:

5

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 24 base pairs

10

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

15

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

20

(A) DESCRIPTION: /desc = "synthetic DNA"

25

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

30

CCSCAGATCA ACAAGTTYTA SGAC

24

35

40

45

50

55

## SEQUENCE LISTING

## (1) GENERAL INFORMATION:

## (i) APPLICANT:

(A) NAME: Japan Science and Technology Corporation  
 (B) STREET: 4-1-8, Honcho  
 (C) CITY: Kawaguchi-shi  
 (D) STATE: Saitama  
 (E) COUNTRY: Japan  
 (F) POSTAL CODE (ZIP): 332-0012  
 (G) TELEPHONE: 81-48-226-5618  
 (H) TELEFAX: 81-48-226-5652

(A) NAME: The Institute of Physical and Chemical Research  
 (B) STREET: 2-1, Hirose  
 (C) CITY: Wako-shi  
 (D) STATE: Saitama  
 (E) COUNTRY: Japan  
 (F) POSTAL CODE (ZIP): 351-0198  
 (G) TELEPHONE: 81-48-462-1111  
 (H) TELEFAX: 81-48-462-4609

(ii) TITLE OF INVENTION: POLYESTER SYNTHASE AND A GENE CODING FOR THE SAME

(iii) NUMBER OF SEQUENCES: 4

## (iv) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk  
 (B) COMPUTER: IBM PC compatible  
 (C) OPERATING SYSTEM: PC-DOS/MS-DOS  
 (D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)

## (vi) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER: JP 82965/1997  
 (B) FILING DATE: 01-APR-1997

## (2) INFORMATION FOR SEQ ID NO: 1:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 559 amino acids  
 (B) TYPE: amino acid  
 (C) STRANDEDNESS:  
 (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

Met Ser Asn Lys Asn Ser Asp Asp Leu Asn Arg Gln Ala Ser Glu Asn  
 1 5 10 15  
 Thr Leu Gly Leu Asn Pro Val Ile Gly Leu Arg Gly Lys Asp Leu Leu  
 20 25 30  
 Thr Ser Ala Arg Met Val Leu Thr Gln Ala Ile Lys Gln Pro Ile His  
 35 40 45  
 Ser Val Lys His Val Ala His Phe Gly Ile Glu Leu Lys Asn Val Met  
 50 55 60 65  
 Phe Gly Lys Ser Lys Leu Gln Pro Glu Ser Asp Asp Arg Arg Phe Asn  
 70 75 80  
 Asp Pro Ala Trp Ser Gln Asn Pro Leu Tyr Lys Arg Tyr Leu Gln Thr  
 85 90 95  
 Tyr Leu Ala Trp Arg Lys Glu Leu His Asp Trp Ile Gly Asn Ser Lys  
 100 105 110  
 Leu Ser Glu Gln Asp Ile Asn Arg Ala His Phe Val Ile Thr Leu Met  
 115 120 125  
 Thr Glu Ala Met Ala Pro Thr Asn Ser Ala Ala Asn Pro Ala Ala Val  
 130 135 140 145  
 Lys Arg Phe Phe Glu Thr Gly Gly Lys Ser Leu Leu Asp Gly Leu Thr  
 150 155 160  
 His Leu Ala Lys Asp Leu Val Asn Asn Gly Gly Met Pro Ser Gln Val  
 165 170 175  
 Asp Met Gly Ala Phe Glu Val Gly Lys Ser Leu Gly Thr Thr Glu Gly  
 180 185 190

Ala Val Val Phe Arg Asn Asp Val Leu Glu Leu Ile Gln Tyr Arg Pro  
 195 200 205  
 Thr Thr Glu Gln Val His Glu Arg Pro Leu Leu Val Val Pro Pro Gln  
 210 215 220 225  
 Ile Asn Lys Phe Tyr Val Phe Asp Leu Ser Pro Asp Lys Ser Leu Ala  
 230 235 240  
 Arg Phe Cys Leu Ser Asn Asn Gln Gln Thr Phe Ile Val Ser Trp Arg  
 245 250 255  
 Asn Pro Thr Lys Ala Gln Arg Glu Trp Gly Leu Ser Thr Tyr Ile Asp  
 260 265 270  
 Ala Leu Lys Glu Ala Val Asp Val Val Ser Ala Ile Thr Gly Ser Lys  
 275 280 285  
 Asp Ile Asn Met Leu Gly Ala Cys Ser Gly Gly Ile Thr Cys Thr Ala  
 290 295 300 305  
 Leu Leu Gly His Tyr Ala Ala Leu Gly Glu Lys Lys Val Asn Ala Leu  
 310 315 320  
 Thr Leu Leu Val Ser Val Leu Asp Thr Thr Leu Asp Ser Gln Val Ala  
 325 330 335  
 Leu Phe Val Asp Glu Lys Thr Leu Glu Ala Ala Lys Arg His Ser Tyr  
 340 345 350  
 Gln Ala Gly Val Leu Glu Gly Arg Asp Met Ala Lys Val Phe Ala Trp  
 355 360 365  
 Met Arg Pro Asn Asp Leu Ile Trp Asn Tyr Trp Val Asn Asn Tyr Leu  
 370 375 380 385  
 Leu Gly Asn Glu Pro Pro Val Phe Asp Ile Leu Phe Trp Asn Asn Asp  
 390 395 400  
 Thr Thr Arg Leu Pro Ala Ala Phe His Gly Asp Leu Ile Glu Met Phe  
 405 410 415  
 Lys Asn Asn Pro Leu Val Arg Ala Asn Ala Leu Glu Val Ser Gly Thr  
 420 425 430  
 Pro Ile Asp Leu Lys Gln Val Thr Ala Asp Ile Tyr Ser Leu Ala Gly  
 435 440 445  
 Thr Asn Asp His Ile Thr Pro Trp Lys Ser Cys Tyr Lys Ser Ala Gln  
 450 455 460 465  
 Leu Phe Gly Gly Lys Val Glu Phe Val Leu Ser Ser Ser Gly His Ile  
 470 475 480  
 Gln Ser Ile Leu Asn Pro Pro Gly Asn Pro Lys Ser Arg Tyr Met Thr  
 485 490 495  
 Ser Thr Asp Met Pro Ala Thr Ala Asn Glu Trp Gln Glu Asn Ser Thr  
 500 505 510  
 Lys His Thr Asp Ser Trp Trp Leu His Trp Gln Ala Trp Gln Ala Glu  
 515 520 525  
 Arg Ser Gly Lys Leu Lys Lys Ser Pro Thr Ser Leu Gly Asn Lys Ala  
 530 535 540 545  
 Tyr Pro Ser Gly Glu Ala Ala Pro Gly Thr Tyr Val His Glu Arg  
 550 555 560

## (2) INFORMATION FOR SEQ ID NO: 2:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1680 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: double  
 (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: DNA (genomic)

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

ATGAGTAACA AGAATAGCGA TGACTTGAAT CGTCAAGCCT CGGAAAACAC CITGGGGCTT 60  
 AACCCGTGCA TCGGCCTGCG TGGAAAAGAT CTGCTGACTT CTGCCCGAAT GGTTTTAACC 120  
 CAAGCCATCA AACAAACCAT TCACAGCGTC AAGCACGTCG CGCAITTTGG CATCGAGCTG 180  
 AAGAACGTGA TGTTTGCAA ATCGAAGCTG CAACCGGAAA GCGATGACCG TCGTTTCAAC 240  
 GACCCCGCCT GGAGTCAGAA CCCACTCTAC AAACGTTATC TACAAACCTA CCTGGCGTGG 300  
 CGCAAGGAAC TCCACGACTG GATCGGCAAC AGCAAACTGT CCGAACAGGA CATCAATCGC 360  
 GCTCACTTCG TGATCACCTT GATGACGAA GCCATGGCCC CGACCAACAG TCGCGCCAAT 420  
 CCGGCGGCGG TCAAACGCTT CTTGAAACC GCGGTAATAA GCCTGCTCGA CGGCCTCACA 480  
 CATCTGGCCA AGGACCTGGT AAACAACGCG GGCATGCCGA GCCAGGTGGA CATGGGCGCT 540  
 TTCCAAGTCG GCAAGAGTCT GGGGACGACT GAAGGTGCAG TGGTTTTCCG CAACGACGTC 600  
 CTCGAATTGA TCCAGTACCG GCCGACCACC GAACAGGTGC ATGAGCGACC GCTGCTGGTG 660  
 GTCCCAACCGC AGATCAACAA GTTTTATGTG TTTGACCTGA GCCCGGATAA AAGCCTGGCG 720

CGCTTCTGCC	TGAGCAACAA	CCAGCAAAAC	TTTATCGTCA	GCTGGCGCAA	CCCAGCAAG	780
GCCCAAGCGTG	AGTGGGCTGT	CGTGACTTAC	ATCGATCGGC	TCAAGAAGC	CGTGCAGTA	800
GTTTTCGCGA	TACCCGGCAG	CAAGAAGTCT	AACATGCTCT	GCGCTGCTC	CGGTGGCAAT	840
ACCTGCACCG	CGCTGCTGGG	TCACTAGGCC	GCTCTCGGGC	AGAAGAAGT	CAATGCCCTG	960
ACCCCTTTTG	TCAGCGTGCT	CGACACCCAC	CTCGACTCCC	AGGTTGCATC	TTTCTGCGAT	1020
GAGAAAACCC	TGGAAGCTGC	CAAGCGTTCAC	TGTTATCAGG	CCGGCGTGCT	GTAAGGGCCG	1080
GACATGGCCA	AGTGTCTTCG	CTGATGCGC	CCTAACGACC	TGATCTGGAA	CTACTGGGTC	1140
AACAACATACC	TCTGGGTTAA	CGGCGCAACG	GTCTTCGACA	TTCTTTCTG	GAAACAAGAC	1200
ACCACCCGGT	TGCCGTGCTG	GTTCCACGGC	GATCTGATCG	AAATGTTCAA	AAATAACCCA	1260
CTGGTGC CGC	CCAAATGCAT	CGAAGTGAGC	GGCAGCCGGA	TGCGACTCAA	ACCTGTGCTC	1320
CGCGACATCT	ATCTCCCTGC	GGCCACCAAC	GATCATACAT	GCCTCGGAA	GCCTTGCTAC	1380
AAGTCGGCGC	AACTGTTCCG	TGGCAAGGTG	GAATTCGTGC	TGTCACGACG	TGGGCATATC	1440
CAGAGCATCC	TGAACCCCGC	GGGCAATCCG	AAATCAGCTT	ACATGACACG	CACCGCATCG	1500
CCAGGCCACCG	CCAAAGCAGT	GCAAGAAAAA	TCAACCAAGC	ACACGCACTC	CTGGTGGGCT	1560
CACCTGGCAGG	CTCTGGCAGC	CGAGCGCTCG	GGCAAACTGA	AAATGTCCTC	CACCGACCTG	1620
GGCAACAAGG	CTTATCCGCT	AGGAAAGACC	CGCCCGGGCA	CGATGTGCA	TGAAGCTTAA	1680

(2) INFORMATION FOR SEO ID NO: 3:

(1) SEQUENCE CHARACTERISTICS:

- SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 1826 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear

(11) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

CTGCAGTGCT	CTCTGAACCTA	GAAAGCAACG	TTGTGCAATT	AACGGTCACC	CGAGCAGTAG	60
TACCTGCGCG	TTGCTGTGTG	ACTACACAGC	TGGTCCGGT	ACTCAAGCTCA	GGCATATTGA	120
CGCTGCTAGA	TGAGTAACAA	TAGATAGCAT	GACTTGTACT	GTCAGTGCTG	GGAAAAACCC	180
TTGGGGCTTA	ACCCTGTCAT	CGGCCTGCGT	GGAAAAAGATC	TGCTGACTTC	TGCCCGAATG	240
GTTTTAAACC	AAGCCATACA	ACAAACGACT	CCACAGCGTCA	AGCAGCTGCG	GCATTTTGGC	300
ATCAGAGCTGA	AGAAAGCTGAT	GTTTGGCAAA	TCGAAGCTGC	AACCCGGAAG	CGAGTACCGT	360
CGTTTCAACG	ACCCCGCCTG	GAGTCAGAAC	CCACTTCTAC	AACGTTATCT	ACAAACCTAC	420
CTGCGCTGGG	GCAAGGAAGT	CCACAGCTGG	ATTGGGCAACA	CGAAACTGTC	CGAACAGGAC	480
ATCAATCGCT	CTCAGTGTGT	GATCACCTCT	ATGACCGAAG	CCATGGCCCC	GACCAACAGT	540
GCGGCCAATC	CGGCGCGCGT	CAAACGCTTC	TTCGAAACCG	GCGGTAAAG	CCTGCTCGAC	600
GGCCTCACAC	ATCTGGCCAA	CTCAGCTGGTA	AACAACCGCG	GAGTGGCCAG	CCAGGTGGAC	660
ATTGGGCGCT	TGCAAGTCGG	CAGAGAGTCG	GGGACGACTG	CGATGTCAGT	GGTTTTCCGC	720
AACGACGTCC	TGCAATTGAT	CCAGTACACG	CGCACACCGC	AACAGGTGCA	TCGCGCAGCC	780
CTGCTGTGTG	TCCCACCGCA	GATCAACAA	TTTTATGTGT	TTGACTGTAG	CGGAGATAAA	840
AGCCCTGGCG	GCTTCTGCCT	GAGCAACAAC	CAGCAAACCT	TTATCGTCAG	CTGGCGCAAC	900
CCGACCAAGG	CCACAGCTGA	GTTGGGCTTG	TGCACTTACA	TGATTCGCGT	CAAGGATGCC	960
GTGCAGCTAG	TTTTCGCCAT	CACCGGCATC	AAGAATCATCA	ACATGCTCGG	CGCTGTGCTG	1020
GGTGGCATT	CCTGCACCGC	GCTGCTGGGT	CACTACGCCG	CTCTGGCGGA	GAAGAAGGTC	1080
AATGGCCATG	CCCTTTTGGT	GGAAGTCTGC	GACACCAACC	TGCACTTCCA	GGTGTGCACT	1140
TTGCTGCATG	AGAAAACCTC	CAGAGCTGTC	AAGCGTCACT	CGTATCAGCG	CGGGTGCTGT	1200
GAAGCGCGCG	ACATGGCCAA	AGTCTTGCC	TGGATGCGCC	CTAACGAACT	GATCTGGAAC	1260
TACTGGGTCA	ACAACTACCT	CTGGGGTACA	GAGCCACCGG	TCTTGCAGAT	TCTTTTCTGG	1320
AACAAGGATA	CCACCGCGTT	GCTCTGTGCG	TTCCACGCCG	ATCTGATCGA	AATGTTTCAA	1380
AATAACCCAC	TGCTGGCGCG	CAATGTCATC	GAAGTGAGCG	GCACGCGCAT	CGACCTCAAA	1440
CAGGTCATCC	CGGACATCTA	CTCCTTGCC	GGCACCAACG	ATCATCATCA	CGCTTGGAA	1500
TCTTGCTACA	AGTGGCGCGA	ACTGTTGGGT	GGCAAGGTG	AATTTGCTGT	GTCCAGCAGT	1560
GGCATATCC	AGAGCACTTC	GAAACGCGCG	GGCAATTCGA	AATCACTGTA	CATGACGACC	1620
CCGCAATATC	GAGCCACCTG	CAACAGGTGT	CAAGAAACTG	CAACCAAGCA	CACCGCACTG	1680
TGGTGGCTGC	ACTGGCAGGC	CTGCGAGGCC	GAGCGCTCGG	GCAAACTGAA	AAAGTCCCCG	1740
ACCAGCGCTG	GCAACAAAGC	CTATTCTGCTA	GGAGAAAGCG	CGCCGGGACG	GTATGTGCAT	1800
GAACGTTAAG	TTGTAGGCGT	CTTAGA	@@	@@	@@	@@

1826

(2) INFORMATION FOR SEQ ID NO: 4:

## (1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 24 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc = "synthetic DNA"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

CCSCAGATCA ACAAGTTYTA SGAC

24

# Claims

1. A polypeptide comprising the amino acid sequence of SEQ ID NO:1 or analogs, variants or fragments thereof.
2. A polyester synthase gene comprising DNA coding for a protein containing the amino acid sequence of SEQ ID NO:1 or an analog, variant or fragment thereof.
3. A polyester synthase gene according to claim 2, wherein the DNA coding for the protein with polyester synthase activity is that of SEQ ID NO:2.
4. A polyester synthase gene comprising the nucleotide sequence of SEQ ID NO:3.
5. A recombinant vector comprising the polyester synthase gene of any one of claims 2 to 4.
6. A transformant transformed with the recombinant vector of claim 4.
7. A process for producing polyester synthase, wherein the transformant of claim 6 is cultured in a medium and polyester synthase is recovered from the resulting culture.





European Patent  
Office

# EUROPEAN SEARCH REPORT

Application Number  
EP 98 30 2554

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.6)
P, X	WO 97 22711 A (REGENTS OF THE UNIVERSITY OF MINNESOTA) 26 June 1997 * page 3, line 10 - page 7, line 31; figures 23J,K; example 3 *	1,2,5-7	C12N15/52 C12N15/60 C12N9/88 C12N15/74 C12N1/21
X	WO 91 00917 A (MASSACHUSETTS INSTITUTE OF TECHNOLOGY) 24 January 1991	1,2,5-7	
Y	* page 5, line 9 - line 30 * * page 7, line 1 - line 5 * * page 36, line 15 - page 40, line 5; figures 5,6 *	3,4	
D, X	TIMM, A. AND STEINBÜCHEL, A.: "Cloning and molecular analysis of the poly(3-hydroxyalkanoic acid) gene locus of Pseudomonas aeruginosa PA01" EUROPEAN JOURNAL OF BIOCHEMISTRY, vol. 209, no. 1, October 1992, pages 15-30, XP002087430	1,2,5-7	
Y	* page 16, left-hand column, line 12 - line 28 * * page 18, left-hand column, line 24 - page 19, right-hand column, line 17 * * page 25 - page 28 * 'Discussion' * figures 1,2 *	3,4	TECHNICAL FIELDS SEARCHED (Int.Cl.6)  C12N
D, X	HUISMAN, G.W. ET AL.: "Metabolism of poly(3-hydroxyalkanoates) (PHAs) by Pseudomonas oleovorans" THE JOURNAL OF BIOLOGICAL CHEMISTRY, vol. 266, no. 4, 5 February 1991, pages 2191-2198, XP002087431	1,2,5-7	
Y	* page 2192 - page 2197 * 'Results' and 'Discussion' * figure 2 *	3,4	
The present search report has been drawn up for all claims			
Place of search MUNICH		Date of completion of the search 10 December 1998	Examiner Donath, C
<p>CATEGORY OF CITED DOCUMENTS</p> <p>X: particularly relevant if taken alone Y: particularly relevant if combined with another document of the same category A: technological background O: non-written disclosure P: intermediate document</p> <p>T: theory or principle underlying the invention E: earlier patent document, but published on, or after the filing date D: document cited in the application L: document cited for other reasons</p> <p>&amp;: member of the same patent family, corresponding document</p>			

EPO FORM 1503 03.92 (P/C01)



European Patent  
Office

# EUROPEAN SEARCH REPORT

Application Number  
EP 98 30 2554

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.6)
Y	<p>TIMM, A. ET AL.: "A general method for identification of polyhydroxyalkanoic acid synthase genes from pseudomonads belonging to the rRNA homology group I"</p> <p>APPLIED MICROBIOLOGY AND BIOTECHNOLOGY, vol. 40, no. 5, January 1994, pages 669-675, XP002087432</p> <p>* page 670 - page 671 *</p> <p>'Identification of PHA synthase genes' and 'Cloning of the PHA synthase genes'</p> <p>* page 673 - page 674 *</p> <p>'Discussion'</p> <p>----</p>	1-7	
Y	<p>STEINBÜCHEL, A. ET AL.: "Molecular basis for biosynthesis and accumulation of polyhydroxyalkanoic acids in bacteria"</p> <p>FEMS MICROBIOLOGY REVIEWS, vol. 103, no. 2-4, December 1992, pages 217-230, XP002087433</p> <p>* page 219 - page 224 *</p> <p>'4. Cloning of PHA-biosynthetic genes'</p> <p>* page 228 - page 229 *</p> <p>'Conclusions'</p> <p>-----</p>	1-7	
The present search report has been drawn up for all claims			TECHNICAL FIELDS SEARCHED (Int.Cl.6)
Place of search		Date of completion of the search	Examiner
MUNICH		10 December 1998	Donath, C
<p><b>CATEGORY OF CITED DOCUMENTS</b></p> <p>X : particularly relevant if taken alone</p> <p>Y : particularly relevant if combined with another document of the same category</p> <p>A : technological background</p> <p>O : non-written disclosure</p> <p>P : intermediate document</p> <p>T : theory or principle underlying the invention</p> <p>E : earlier patent document, but published on, or after the filing date</p> <p>D : document cited in the application</p> <p>L : document cited for other reasons</p> <p>.....</p> <p>&amp; : member of the same patent family, corresponding document</p>			

EPO FORM 1503 03/82 (P44/01)

**ANNEX TO THE EUROPEAN SEARCH REPORT  
ON EUROPEAN PATENT APPLICATION NO.**

EP 98 30 2554

This annex lists the patent family members relating to the patent documents cited in the above-mentioned European search report. The members are as contained in the European Patent Office EDP file on  
The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

10-12-1998

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9722711 A	26-06-1997	EP 0870053 A	14-10-1998
WO 9100917 A	24-01-1991	AT 172497 T	15-11-1998
		CA 2062816 A	11-01-1991
		DE 69032713 D	26-11-1998
		EP 0482077 A	29-04-1992
		EP 0870837 A	14-10-1998
		JP 5500751 T	18-02-1993
		US 5480794 A	02-01-1996
		US 5534432 A	09-07-1996
		US 5663063 A	02-09-1997
		US 5245023 A	14-09-1993
		US 5250430 A	05-10-1993

EPO FORM P469

For more details about this annex : see Official Journal of the European Patent Office, No. 12/82